Environmental risk assessment of endocrine disrupters: status and needs of biomarkers



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Key Principle of ERA (aquatic)

Predicted Environmental Concentration (PEC)

Predicted No Effect Concentration (PNEC)

PEC:PNEC ratio

Calculation typically includes data on:

- Microorganisms (from Sewage Treatment Works)
- Plants
- Invertebrates
- Fish

Finally, if the PEC:PNEC ratio \geq 1, potential risk and further work (termed 'risk refinement') may be required.



Biomarker definition

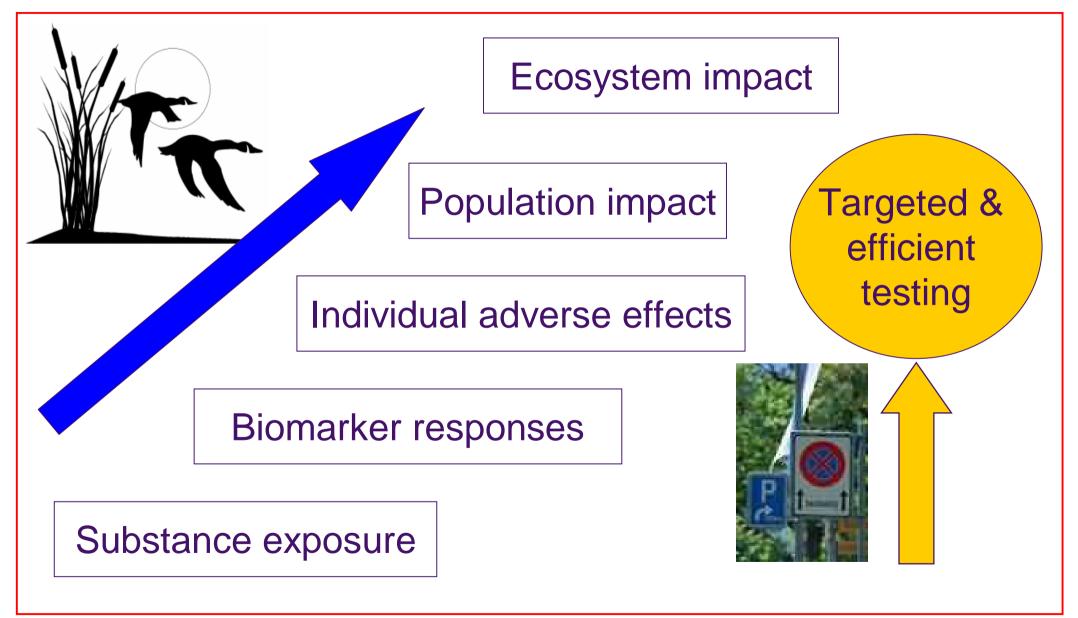
"A molecular, biochemical, cellular or physiological indicator of either exposure to, or effects of, xenobiotic chemicals".

> (after Huggett *et al.* (1992) Biomarkers – SETAC Special Publication.

In this context, biomarker responses <u>do not</u> <u>include</u> commonly measured toxicological effects such as reduced development, fecundity, fertilisation, growth or survival.



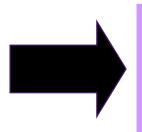
'Biomarkers as signposts ...' concept





Three key issues to consider ...

- Which biomarkers are available today for measuring endocrine disruption in field and laboratory studies?
- Have the biomarkers of interest been adequately validated in different laboratories?
- From a science perspective, how should biomarkers be used in regulatory testing to support environmental risk assessment (ERA)?



Major goals: better understanding, reliable measurements & accurate predictions.

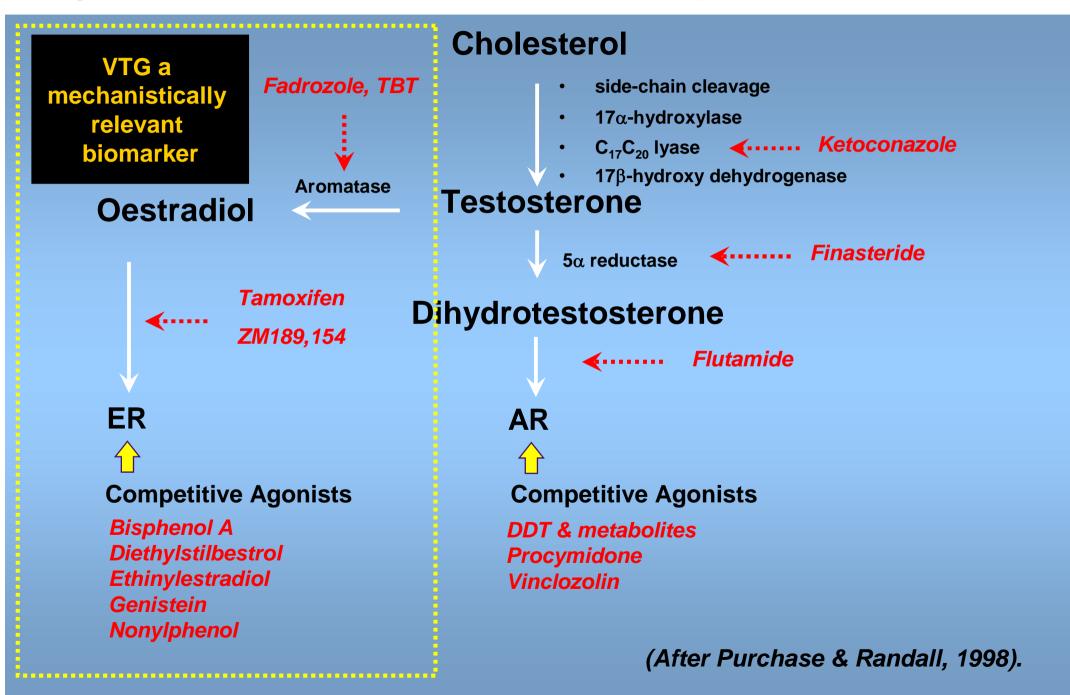
1. Which biomarkers are available today for EDCs?

- Endocrine system biomarkers
 - gene expression (eg AR, ER, CYP19)
 - plasma hormone levels (eg oestradiol, thyroxine)
 - vitellogenin
 - secondary sexual characteristics
 - gonad histology
- In vitro applications
 - vertebrate cell lines (eg fish hepatocytes)
 - invertebrate cell lines (eg insect Bll cells)
- In vivo applications
 - field studies
 - laboratory testing

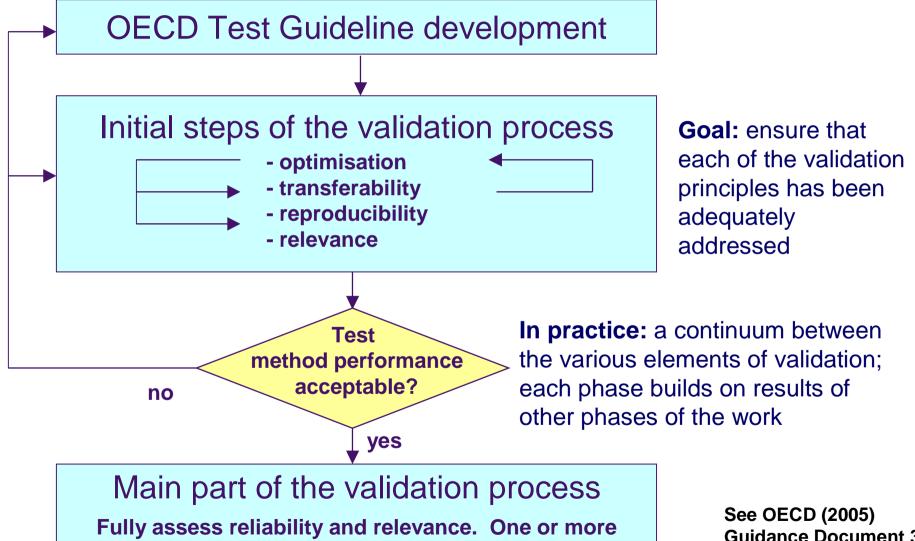
Refs: Fent (2001) Toxicol In Vitro 15: 477–488 (fish cell lines); Dinan et al (2001) Env.Toxicol.Chem. 20: 2038– 2046 (insect cell line); Hutchinson et al (2006) Environ.Health Perspect. – in press (fish biomarkers review)

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Key mechanisms – examples



2. Have the biomarkers of interest been adequately validated?



phases including inter-laboratory testing, blind testing and assessment of positive/negative controls **Guidance Document 34.**



Validation terminology

- Repeatability = the variation between repeated tests of the same protocol in the same laboratory. This is also caused intra-laboratory variability.
- Reproducibility = the variation between repeated tests of the same protocol in different laboratories. This is also caused interlaboratory variability.
- Comparability = the variation for the same biological endpoint measured using different protocols

Refs: Hutchinson et al (2006) Environ. Health Perspect. – in press (fish biomarkers review)



Case study - vitellogenin

Organism	Endpoint	Variable	CV (%)
Fathead minnow	Juvenile whole body VTG	Reproducibility	55
67	Female plasma VTG	Reproducibility	45
	Male plasma VTG	Reproducibility	38
s 7	Plasma VTG	Repeatability	3 - 14
s 7	Plasma VTG	Repeatability	16.4
5.7	Plasma VTG	Reproducibility	18.6
4.7	Plasma VTG	Repeatability	32
• ?	Plasma testosterone	Repeatability	46
17	Plasma estradiol	Repeatability	64
Medaka	Liver VTG	Comparability	52 - 863
	Whole body VTG	. ,	100 - 1873
	Liver VTG	Repeatability	<7
Zebrafish	Whole body VTG	Comparability	70.2 - 269
	Whole body VTG	Repeatability	14 - 18
Medaka	Fecundity	Repeatability	35.7
Rainbow trout	28d growth LOEC	Repeatability	_
47	28d growth LOEC	Reproducibility	19 - 58
Sheepshead minnow	Larval IC25	Repeatability	28 - 42
67	Larval IC25	Reproducibility	44
Zebrafish	Survival NOEC	Repeatability	26 - 33
67	Survival NOEC	Reproducibility	35 - 52
	Fecundity	Repeatability	26 - 63

From: Hutchinson et al (2006) Environ. Health Perspect. – in press (fish biomarkers review).



Validation – overall situation

- Vitellogenin probably the leading example:
 - mechanistic scope is well understood
 - repeatability well-defined
 - reproducibility looks as good as traditional ecotoxicology endpoints (19-55%)
- BUT more work is needed on other biomarkers:
 - plasma sex steroid levels
 - plasma thyroid hormones
 - gonad histology
 - molecular responses
- Historical databases are essential tools for interpretation of biomarker data*

Lesson number 2 from Sumpter and Johnson (2005) Environ. Sci. Technol. 39: 4321-4332

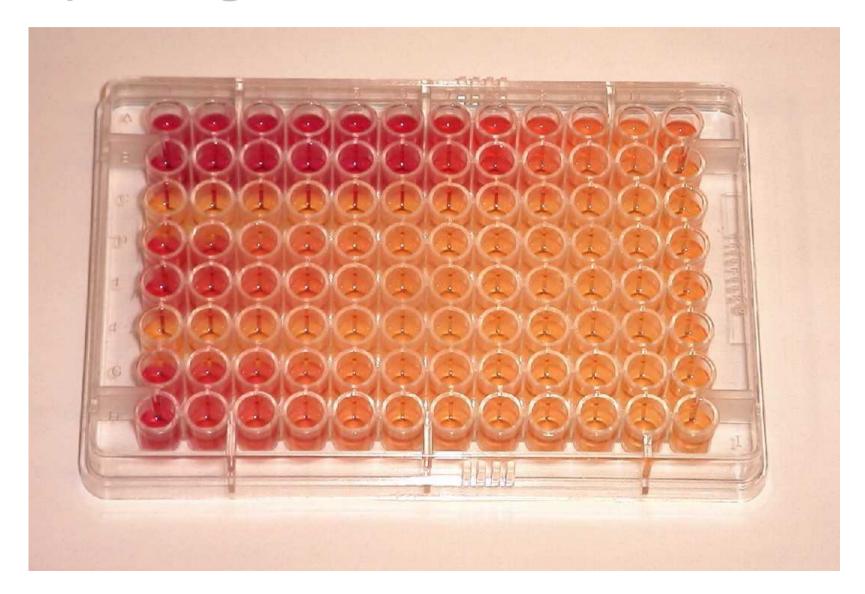


3. How should biomarkers be used in regulatory testing?

- Validated biomarkers are important in key areas:
 - **1. Linking field and laboratory studies**
 - 2. Screening for endocrine specific mechanisms of action
 - 3. Helping efficient design of chronic tests for risk assessment
 - species selection
 - exposure concentration selection



YES and YAS for (anti-)oestrogenic and (anti-)androgenic activities





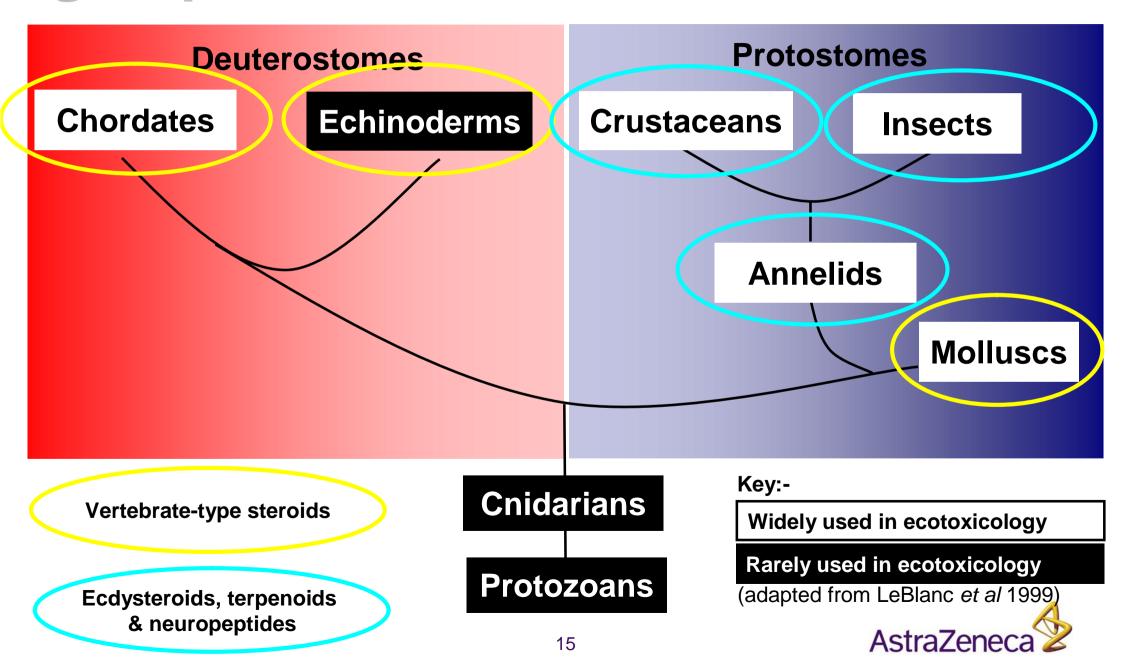
In vitro BII Assay for Ecdysteroid Activity

<u>Substance</u>	Concn	<u>Response (+/-)</u>
Bisphenol A	3.0 x 10-3M	Active antagonist
DES	1.0 x 10⁻³M	Inactive
Diethylphthalate	2.0 x 10-3M	Active antagonist
E2	3.0 x 10⁻⁵M	Inactive
Flutamide	5.0 x 10 ⁻⁵ M	Inactive
Genistein	1.0 x 10⁻³M	Inactive
20-hydroxyecdysone	7.6 x 10 ⁻⁹ M (~4 ppb)	Active agonist
Lindane	3.0 x 10-3M	Active antagonist
Methoxychlor	1.0 x 10⁻³M	Inactive
Octylphenol	1.0 x 10 ⁻³ M	Inactive

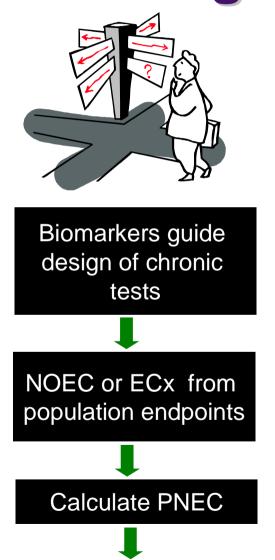
Dinan et al (2001) Environ Toxicol Chem 20: 2038-2046



Phylogeny of major invertebrate groups within Coelomates

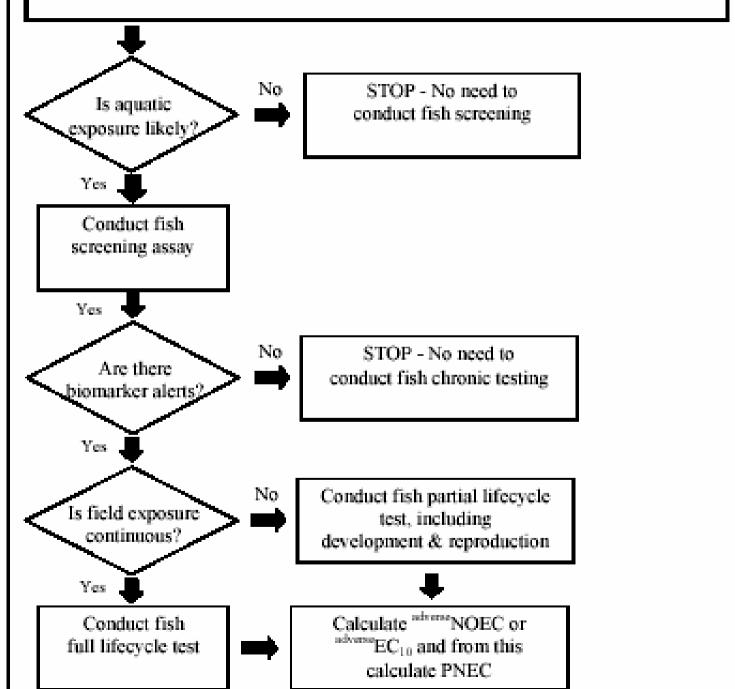


Biomarkers for fish screening

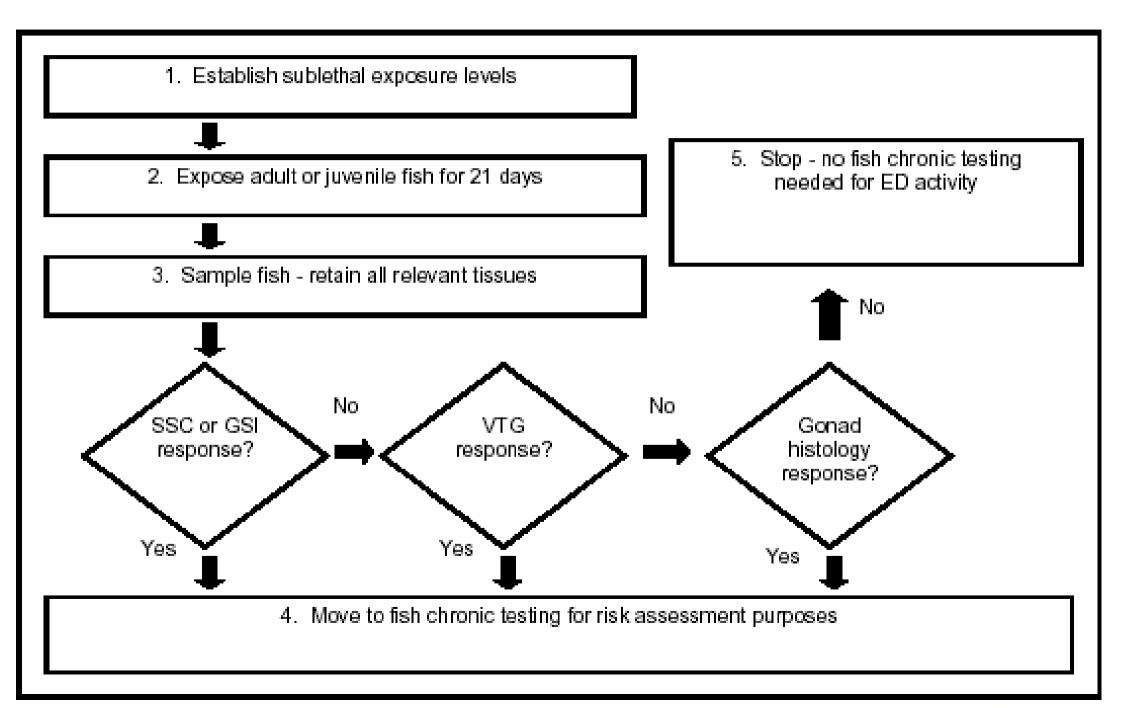


Biomarkers to link lab - field Review existing information:

- in vitro or in silico data
- alerts based on chemical structure
- alerts from mammalian, avian or amphibian data



Fish screening – stepwise biomarkers



Biomarker responses are not adverse effects

 biomarker EC₁₀, NOEC or LOEC VTG SSC GSI Gonad histology Plasma steroids 	 adverse EC₁₀, NOEC or LOEC survival length weight development fecundity
 Plasma steroids Enzyme induction Gene expression 	fecundityfertilisation ratehatching success

Biomarker signals provide mechanistic data to guide chronic testing for adverse effects. At present they should not be used to derive PNEC values for EDCs.



Adverse effect measures address population relevant parameters and should currently be used for Calculating PNEC values for EDCs.

Acknowledgments

 Thanks to Gary Ankley (US Environmental Protection Agency, Minnesota), Helmut Segner (University of Bern, Switzerland) and Charles Tyler (University of Exeter, UK) for contributing to these ideas.

• For further information see article (available on-

Ine): Hutchinson TH, Ankley GT, Segner H, Tyler CR (2006). Screening and testing for endocrine disruption in fish - biomarkers as signposts not traffic lights in risk assessment. *Environ Health Perspect*

Thank you for your attention!

